

Atmospheric Pressure Matrix Assisted Laser Desorption/Ionization (AP-MALDI) Combined with Ion Trap Mass Spectrometer (ITMS) – a New Technique for Fingerprinting and Structural Analysis of Plant Derived Oligosaccharides



Sun-Li Chong¹, Teemu Nissila², Raimo Ketola², Sanna Koutaniemi¹, Maija Tenkanen¹, Päivi Tuomainen¹

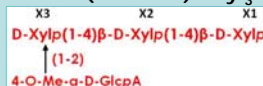
¹Department of Applied Chemistry and Microbiology, Faculty of Agriculture and Forestry, P.O. Box 27, FI-00014 University of Helsinki, Finland. ²Centre for Drug Research, Faculty of Pharmacy, P.O. Box 56, FI-00014 University of Helsinki, Finland

Introduction:

Xylans are the most abundant hemicellulose present in plant secondary cell wall. Most of the xylan structures hitherto characterised were non-acetylated due to alkaline extraction used as the isolation method. Intact xylans are needed for the structural studies in biosynthesis research in order to gain complete understanding of plant behaviour, especially the study of acetylation mechanism in xylan biosynthesis. Thus, a more refined method which circumvents deacetylation or destruction of glucuronic acid groups during xylan isolation is needed for biosynthesis research. Degradation by microbial enzymes is a gentle method and their activities are substrate specific and therefore can be used for fingerprinting the xylan structures in combination with mass spectrometric (MS) detection. We are aiming to develop a MS method that can profile the endoxylanase hydrolysed xylo-oligosaccharides (XOS) (both acetylated and glucuronic acid linked) and perform structural analysis of the hydrolysed products. AP-MALDI in combination with an ion trap mass spectrometer (ITMS) is an interesting system due to its ability to study the degree of polymerization and structures of oligosaccharides. Therefore, we will test the suitability of this system against various model samples and compare the performance with vacuum MALDI-TOF.

Methodology and Results

1) Alkaline extracted birch glucuronoxylan (non-acetylated) was exhaustively hydrolysed by commercially available glycoside hydrolase (GH) family 10 endoxylanase (Shearzyme). The main resulting acidic XOS is (MeGlcA)³-Xyl₃



Hydrolysates were desalted using Dowex 50W(H+)

AP-Maldi-ITMS

- Nitrogen Laser source: 337nm; mass range: standard (*m/z* 50-2000; extended: up to *m/z* 4000)
- Calibration: ESI tuning mix; custom made acidic XOS (UXX), *m/z* 627.17

Performance of both systems were compared

Vacuum Maldi-TOF

- Laser source: 337 nm, 50 Hz; acceleration voltage: 25kV; 1000-2000 shoots were averaged.
- Calibration: Manufacturer supplied protein standard (*m/z*1000-3000); custom made acidic XOS (UXX), *m/z* 627.17

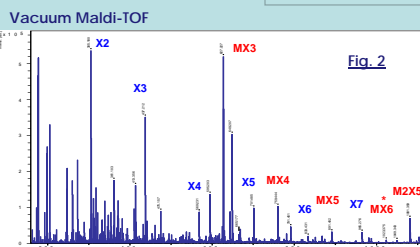
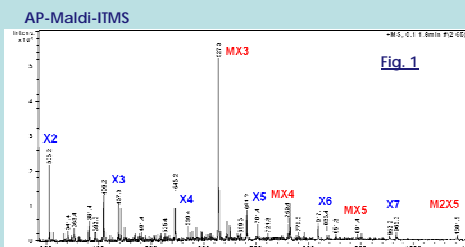


Fig. 1 & 2: Hydrolysates were analysed in positive AP-Maldi-ITMS and vacuum Maldi-TOF. Sodium adducted ions were observed in both spectra.

Both systems were able to detect the major neutral and acidic xylo-oligosaccharides derived from Shearzyme hydrolysed birch xylan. Except 4-O-methyl-glucuronic acid alpha 1-2 linked xylohexaose (MX6, mark with *) was undetectable by AP-Maldi-ITMS.

Xn: Xylo-oligosaccharides, m = 2, 3, 4,
MXn: 4-O-Methyl-Glucuronic acid 1-2 alpha linked XOS, n = 3, 4, 5,

2) Acetylated Xylo-oligosaccharides

Steam exploded eucalyptus XOS were separated into neutral and acidic fractions using porous graphitised carbon (PGC) column and analysed in AP-Maldi-ITMS.

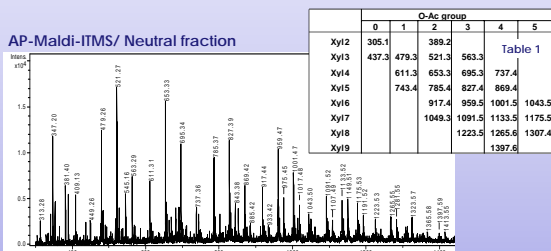


Fig. 3: AP-Maldi was able to detect acetylated XOS up to DP 9. The peaks were identified and listed in Table 1.

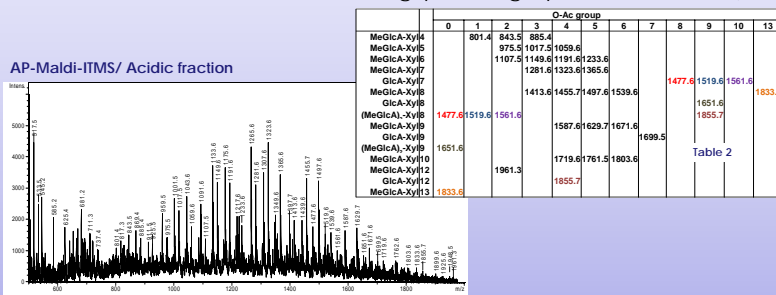


Fig. 4: AP-Maldi was able to detect acidic acetylated XOS up to DP 13. The peaks were identified as 4-O-Methyl-glucuronic acid (MeGlcA) and glucuronic acid (GlcA) alpha 1-2 linked XOS containing different degree of acetylation. The peaks were identified and listed in Table 2. Some overlapped peaks were found and highlighted in same colour.

3) Structural Analysis of Two Xylo-oligosaccharides Isomers

Two aldotetrauronic acid XOS isomers, 4-O-methyl-glucuronic acid 1-2 alpha linked xylotriase, (MeGlcA)³-Xyl₃ and (MeGlcA)²-Xyl₃ were fragmented by ion trap collision induced dissociation.

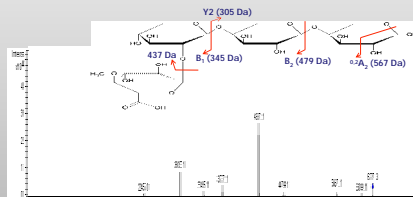


Fig. 5: Positive AP-Maldi-IT- CID mass spectra of (MeGlcA)³-Xyl₃.

The fragment ions were assigned according to fragmentation nomenclature introduced by Doman B and Costello CE. A systematic nomenclature for carbohydrate fragmentations in FAB MS/MS Spectra of glycoconjugates. Glycoconjugate J. 5 (1988) 397-405.

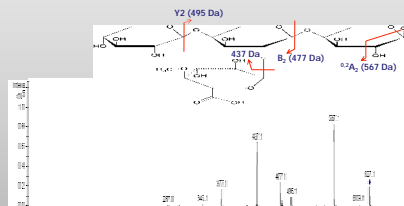


Fig. 6: Positive AP-Maldi-IT- CID mass spectra of (MeGlcA)²-Xyl₃. The MS2 experiment have shown different fragmentation pattern for both isomers.

As a conclusion, AP-MALDI-ITMS was shown to be a potential technique and can be a method of choice for the fingerprinting and structural analysis of plant oligosaccharides in addition to vacuum MALDI-TOF.

